



Dynamics of a methanol-fed marine denitrifying biofilm: 2—impact of environmental changes on the microbial community

Richard Villemur¹, Geneviève Payette¹, Valérie Geoffroy², Florian Mauffrey³ and Christine Martineau⁴

¹ INRS-Centre Armand-Frappier Santé et Biotechnologie, Laval, Québec, Canada

² Lallemand, Montréal, Québec, Canada

³ Université de Genève, Geneva, Switzerland

⁴ Laurentian Forestry Centre, Québec, Canada

ABSTRACT

Background. The biofilm of a methanol-fed, marine denitrification system is composed of a multi-species microbial community, among which *Hyphomicrobium nitratorans* and *Methylophaga nitratorans* are the principal bacteria involved in the denitrifying activities. To assess its resilience to environmental changes, the biofilm was cultivated in artificial seawater (ASW) under anoxic conditions and exposed to a range of specific environmental conditions. We previously reported the impact of these changes on the denitrifying activities and the co-occurrence of *H. nitratorans* strain NL23 and *M. nitratorans* in the biofilm cultures. Here, we report the impact of these changes on the dynamics of the overall microbial community of the denitrifying biofilm.

Methods. The original biofilm (OB) taken from the denitrification system was cultivated in ASW under anoxic conditions with a range of NaCl concentrations, and with four combinations of nitrate/methanol concentrations and temperatures. The OB was also cultivated in the commercial Instant Ocean seawater (IO). The bacterial diversity of the biofilm cultures and the OB was determined by 16S ribosomal RNA gene sequences. Culture approach was used to isolate other denitrifying bacteria from the biofilm cultures. The metatranscriptomes of selected biofilm cultures were derived, along with the transcriptomes of planktonic pure cultures of *H. nitratorans* strain NL23 and *M. nitratorans* strain GP59.

Results. High proportions of *M. nitratorans* occurred in the biofilm cultures. *H. nitratorans* strain NL23 was found in high proportion in the OB, but was absent in the biofilm cultures cultivated in the ASW medium at 2.75% NaCl. It was found however in low proportions in the biofilm cultures cultivated in the ASW medium at 0–1% NaCl and in the IO biofilm cultures. Denitrifying bacterial isolates affiliated to *Marinobacter* spp. and *Paracoccus* spp. were isolated. Up regulation of the denitrification genes of strains GP59 and NL23 occurred in the biofilm cultures compared to the planktonic pure cultures. Denitrifying bacteria affiliated to the *Stappia* spp. were metabolically active in the biofilm cultures.

Conclusions. These results illustrate the dynamics of the microbial community in the denitrifying biofilm cultures in adapting to different environmental conditions. The

Submitted 17 April 2019
Accepted 12 July 2019
Published 13 August 2019

Corresponding author
Richard Villemur,
richard.villemur@iaf.inrs.ca

Academic editor
Joseph Gillespie

Additional Information and
Declarations can be found on
page 26

DOI 10.7717/peerj.7467

© Copyright
2019 Villemur et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

NaCl concentration is an important factor affecting the microbial community in the biofilm cultures. Up regulation of the denitrification genes of *M. nitratreducentescens* strain GP59 and *H. nitratorans* strain NL23 in the biofilm cultures suggests different mechanisms of regulation of the denitrification pathway in the biofilm. Other denitrifying heterotrophic bacteria are present in low proportions, suggesting that the biofilm has the potential to adapt to heterotrophic, non-methylotrophic environments.

Subjects Biodiversity, Biotechnology, Marine Biology, Microbiology

Keywords Denitrification, Biofilm, *Hyphomicrobium*, *Methylophaga*, Marine environment, Metatranscriptome, Microbial diversity

INTRODUCTION

Most naturally-occurring microbial biofilms, such as those encountered in bioremediation processes, are composed of multiple microbial species. Studying such complex biofilms is a challenge, as each species can influence the biofilm development. The biofilm microbial community inside a bioremediation process adapts to the prescribed operating conditions and shapes the efficiency of the bioprocess to degrade the pollutant(s). Usually, the microbial community in such bioprocesses is complex and composed of main degraders but also of secondary microorganisms that could provide benefits to the degraders or could simply contribute to the degradation intermediates or waste. It is recognized that a complex microbial community is more resilient to “unexpected” changes in the operation of the bioprocesses than a single species biofilm, as some of the minor degraders take over the main degraders affected by the changes (*Cabrol & Malhautier, 2011; Roder, Sorensen & Burmole, 2016; Salta et al., 2013; Tan et al., 2017*). The mechanisms of how a microbial population in a biofilm adapts to changes, however, are poorly understood.

The Montreal Biodome, a natural science museum, operated a continuous fluidized-bed methanol-fed denitrification reactor to remove nitrate (NO_3^-) that accumulated in the 3 million-L seawater aquarium. The fluidized carriers in the denitrification reactor were colonized by naturally-occurring multispecies microorganisms to generate a marine methylotrophic denitrifying biofilm estimated to be composed of 15–20 bacterial species (*Labbé et al., 2003*). The main bacteria responsible of the denitrifying activities belong to the alphaproteobacteria *Hyphomicrobium nitratorans* (strain representative NL23) and to the gammaproteobacteria *Methylophaga nitratreducentescens* (strain representative JAM1), both methylotrophs, that accounted for 60–80% of the biofilm (*Labbé et al., 2003; Labbé et al., 2007; Martineau et al., 2013b; Villeneuve et al., 2013*).

Denitrification takes place in bacterial cells where N oxides serve as terminal electron acceptors instead of oxygen (O_2) for energy production when oxygen depletion occurs, leading to the production of gaseous nitrogen (N_2). Four sequential reactions are strictly required for the reduction of NO_3^- to gaseous nitrogen, via nitrite (NO_2^-), nitric oxide (NO) and nitrous oxide (N_2O), and each of these reactions is catalyzed by different enzymes, namely NO_3^- reductases (Nar and Nap), NO_2^- reductases (NirS and NirK), NO reductases (Nor) and N_2O reductases (Nos) (*Kraft, Strous & Tegetmeyer, 2011; Philippot*

et al., 1999; Richardson et al., 2001). Whereas *H. nitrativorans* strain NL23 possesses the four reductases for the complete denitrification pathway, *M. nitratireducentescens* strain JAM1 performs incomplete denitrifying activities, as it lacks a dissimilatory NO-forming nitrite reductase (Auclair et al., 2010; Martineau et al., 2013a; Mauffrey et al., 2017; Mauffrey, Martineau & Villemur, 2015; Villeneuve et al., 2012). Using degenerated PCR primers for the detection of denitrification genes, we showed that there are probably other denitrifying bacteria in the biofilm, one to four orders of magnitude lower in proportions than *M. nitratireducentescens* strain JAM1 and *H. nitrativorans* strain NL23 (Auclair, Parent & Villemur, 2012). These other bacteria may play a role if the bioprocess undergoes stress conditions or changes in the operation mode.

We have initiated a study with the aim of assessing the performance of the Biodome denitrifying biofilm subjected to environmental changes. The original biofilm (OB) taken from the Biodome denitrification system was cultivated in an artificial seawater (ASW) under batch mode and anoxic conditions at laboratory scale and exposed to a range of specific physico-chemical parameters. Such parameters included a range of NaCl, NO_3^- , NO_2^- and methanol concentrations, and varying pH and temperature. These parameters were chosen as possible factors that could affect a denitrification reactor. Thus, the objectives of this study were to determine the impact of these changes: (1) on the denitrification performance of the biofilm; (2) on the dynamics of the co-occurrence of *H. nitrativorans* and *M. nitratireducentescens* in the biofilm; and (3) on the overall microbial community. The fourth objective of the study was to determine whether denitrifying bacteria other than *H. nitrativorans* strain NL23 and *M. nitratireducentescens* strain JAM1 are present in the biofilm.

Results for the first two objectives and partially the fourth objective were reported by Geoffroy et al. (2018) and Payette et al. (2019). We showed that the denitrifying biofilm can sustain denitrifying activities in most of the tested conditions. Inhibition occurred when these biofilm cultures were exposed to high pH (10) or to high methanol concentrations (1.5%). The highest specific denitrification rates occurred when the biofilm cultures were cultivated at 64.3 mM NO_3^- and 0.45% methanol (C/N = 1.5), and at 30 ° C. Poor biofilm development occurred in biofilm cultures cultivated at 5% and 8% NaCl. We also showed that the NaCl concentrations in the ASW medium have significant impacts on the population of *H. nitrativorans* strain NL23, with its displacement by a subpopulation of the species *M. nitratireducentescens* (strain GP59 as representative), which can perform the complete denitrification pathway.

Results for the third and fourth objectives are presented here. The composition of the bacterial community of the different biofilm cultures was determined by sequencing the 16S ribosomal RNA (rRNA) genes. A culture dependent approach was used to recover new denitrifying bacterial isolates from the biofilm cultures. To complement these two objectives, we derived the metatranscriptome from selected biofilm cultures. These metatranscriptomes were analyzed to determine the composition of the active microbial community in the biofilm cultures but also to assess their metabolic contributions, such as those involved in denitrification. Furthermore, metatranscriptomic analyses provided further indications on the dynamics of *H. nitrativorans* and *M. nitratireducentescens*

Table 1 Biofilm culture conditions.

Name	Medium	NO ₃ ⁻ mM (mg-NO ₃ -N/L) ^c	Methanol % (v/v) ^c	NaCl % (w/v)	Temp °C	Specific ^a denitrification rates mM-NO _x h ⁻¹ mg-protein ⁻¹
Ref300N-23C ^b	ASW	21.4 (300)	0.15	2.75	23	0.0530
300N-30C	ASW	21.4 (300)	0.15	2.75	30	0.0946
900N-23C	ASW	64.3 (900)	0.45	2.75	23	0.0637
900N-30C	ASW	64.3 (900)	0.45	2.75	30	0.0979
0%NaCl	ASW	21.4 (300)	0.15	0	23	0.0911
0.5%NaCl	ASW	21.4 (300)	0.15	0.5	23	0.0712
1%NaCl	ASW	21.4 (300)	0.15	1.0	23	0.0357
IO	IO	21.4 (300)	0.15	3.0 ^d	23	0.0611

Notes.

The original biofilm was cultured in triplicates in these conditions at pH 8.0. The carriers were transferred 5 times in fresh medium around each week before measuring the denitrifying activities.

^aFrom Payette et al. (2019).

^bReference biofilm cultures.

^cThe C/N ratio was 1.5 in all biofilm cultures.

^dThe exact amount of NaCl added in the IO medium is not known. See Payette et al. (2019) for the IO composition. For comparison, the amount of Na⁺ and Cl⁻ in the ASW medium is 3.2%.

In gray are changed parameters from the reference biofilm cultures.

IO, Instant Ocean medium.

in these cultures (second objective) by assessing changes in metabolic pathways of *H. nitrivorans* strain NL23 and *M. nitratreducentescens* strain GP59 between the planktonic pure cultures and the biofilm cultures. Our study is the first that give a comprehensive picture of the microbial community of a methylotrophic denitrifying biofilm and its adaptation to specific changes.

MATERIAL AND METHODS

Cultivation of the original biofilm to different culture conditions

The formulations of the artificial seawater (ASW) medium and the commercial Instant Ocean (IO) medium (Table S1), and the different conditions of the biofilm cultures were described by Payette et al. (2019). Briefly, the biomass of several carriers taken from the denitrification reactor of the Montreal Biodome was scraped, dispersed, then distributed to several vials containing twenty free carriers and 60 mL prescribed medium (Table 1; Fig. S1). The vials were incubated under anoxic conditions at 23 °C or 30 °C (Table 1) and shaken at 100 rpm (orbital shaker). In average once a week, the twenty carriers were taken, gently washed to remove the excess medium and the planktonic bacteria, then transferred into fresh anoxic medium and incubated under the same conditions (Fig. S1). The Ref300N-23C biofilm cultures (for 300 mg NO₃⁻-N/L, 23 °C) were defined as the *reference biofilm cultures*. These cultures were used by Payette et al. (2019) as a reference to compare results between the different culture conditions. The protocols to measure NO₃⁻ and NO₂⁻ concentrations, and to extract DNA from the biofilm cultures or the planktonic pure cultures were described in Payette et al. (2019) and Geoffroy et al. (2018).

16S rRNA gene analysis

DNA extracted from triplicate biofilm cultures was pooled before sequencing. Total DNA samples from seven biofilm cultures (Table 1; Ref300N-23C, 300N-30C, 900N-23C, 900N-30C, 0%NaCl, 0.5%NaCl and 1.0%NaCl) were sent to the sequencing service at the Research and Testing Laboratory (RTL, Lubbock, Texas, USA). A region of the 16S rRNA gene was PCR amplified using the 28F-519R primers (5' GAGTTTGATCNTGGCTCAG 3' and 5' GTNTTACNGCGGCKGCTG 3', covering the V1–V2–V3 variable regions) and subjected to pyrosequencing using a Roche 454 FLX genome sequencer system. The sequencing service (RTL) performed denoising and chimera analyses (details provided in Document S1). The high-quality reads were then processed in the RDP pipeline at the Ribosomal data project (RDP) web site (Cole *et al.*, 2014). Reads were clustered into operational taxonomic units (OTU) using a 97% identity threshold. DNA extracted from the OB (from frozen stock) and from fresh IO biofilm cultures (Table 1) were sent to the sequencing service of Genome Quebec Innovation Center (Montreal, QC, Canada). In these cases, the 16S rRNA sequences covering the V6–V7–V8 variable regions (5' ACACTGACGACATGGTTCTACA 3' and 5' TACGGTAGCAGAGACTTGGTCT 3') were PCR amplified and sequenced by Illumina MiSeq PE250 (250 bp paired-end sequencing reactions). The reads were processed based on Peck *et al.* (2016). Briefly, paired-end reads were merged with minimum and maximum overlap length between the two reads of 20 and 250 bases, respectively, with 30% mismatched bp tolerance in the overlap region. The merged reads were processed using the software UPARSE (Edgar, 2013). Sequences were truncated to a uniformized length to 420 bp. Reads with a low-quality score were removed using 2.0 as the maximum expected error value. The high-quality reads were de-replicated, sorted by size and singletons were removed. The resulting reads were clustered into operational taxonomic units (OTU) with the UPARSE-OTU clustering method using a 97% identity threshold. Chimeric OTU were removed by UPARSE-REF algorithm and with the software UCHIME ran against ChimeraSlayer 'gold' reference database (Edgar *et al.*, 2011). All representative sequences of the OTUs (from pyrosequencing and Illumina) were checked again for chimeras with the DECIPHER v 2.0 program (<http://www2.decipher.codes/FindChimeras.html>) (Wright, Yilmaz & Noguera, 2012). The affiliation of the OTUs to the most probable genus was determined by the CLASSIFIER program at the RDP web site (Document S2). 16S rRNA sequence reads were deposited in the GenBank Sequence Read Archive (SRA) under the accession number PRJNA524642. Principal component analysis of the proportion of reads associated to the bacterial profiles was performed at ClustVis web site (<https://biit.cs.ut.ee/clustvis/>) (Metsalu & Vilo, 2015).

Isolation of bacterial isolates

Biofilm of the Ref300C-23C biofilm cultures was scraped from the carriers and dispersed in saline solution (3% NaCl, 34.2 mM phosphate buffer pH 7.4), and serial dilutions were made and inoculated onto these agar plate media: (1) R2A medium (complex organic carbons; EMD Chemicals Inc., Gibbstown, NJ, USA), (2) Marine Agar 2216 (marine medium with yeast extract and peptone as carbon source; Becton, Dickinson and Co., Sparks, MD, USA), (3) *Methylophaga* medium 1403 (American Type Culture Collection

[ATCC], Manassas, VA, USA) and (4) the ASW medium; these two latter media were supplemented with 1.5% agar and 0.3% v/v methanol. The isolation procedure, the taxonomic affiliation of the isolates and the measurement of their denitrifying activities were carried out as described by [Geoffroy et al. \(2018\)](#). The 16S rRNA gene sequences were deposited in GenBank under the accession numbers [MK571459–MK571476](#).

Transcriptomes

Planktonic pure cultures of *M. nitratireducenticrescens* strains JAM1 and GP59 were performed in the *Methylophaga* 1403 medium and of *H. nitrativorans* strain NL23 in the 337a medium as described by [Martineau, Mauffrey & Villemur \(2015\)](#) and [Mauffrey, Martineau & Villemur \(2015\)](#). These cultures were carried out in triplicate with methanol (0.3%) and NO_3^- (21.4 mM [300 mg-N/L]) under anoxic conditions at 30 °C. The biomass was collected by centrifugation when the NO_3^- reduction was near completion, and total RNA was extracted as described by [Mauffrey, Martineau & Villemur \(2015\)](#). For the biofilm cultures, at the end of the fifth transfer cultures, the biomass of each replicate was scraped from carriers and used to extract total RNA. The RNA samples were sent to the sequencing service for RNA sequencing (RNAseq) by Illumina (Genome Quebec Innovation Center, Montreal QC, Canada). Because of limited amount of biofilm available, total RNA from the triplicate biofilm samples were pooled before sending to the sequencing service. For the planktonic pure cultures, RNAseq was performed on each replicate. The Ribo-ZeroTM rRNA Removal Kit (Meta-Bacteria; Epicentre, Madison, WI, USA) was used to deplete total RNA of the ribosomal RNA. The RNA was then treated with the TruSeq Stranded mRNA Sample Prep Kit (Illumina Inc, San Diego, CA, USA).

All computations were made on the supercomputer Briarée from the Université de Montréal, managed by Calcul Québec and Compute Canada. Raw reads were filtered to remove low quality reads using FASTX toolkit (http://hannonlab.cshl.edu/fastx_toolkit/) by discarding any reads with more than 10% nucleotides with a PHRED score <20. The resulting reads from each sample/replicate were aligned respectively to the genome of *M. nitratireducenticrescens* strain JAM1 (GenBank accession number [CP003390.3](#)), to the genome and plasmids of *M. nitratireducenticrescens* strain GP59 ([CP021973.1](#), [CP021974.1](#), [CP021975.1](#)) and to the genome of *H. nitrativorans* strain NL23 ([CP006912.1](#)) using Bowtie (v 2.2.3) with default parameters. SAMtools (v 0.1.18) and BEDtools (v 2.20.1) were used for the generation of sam and bam files, respectively. Significance for difference in the relative transcript levels of a gene (defined as transcript per million: TPM) between planktonic pure cultures and biofilm cultures was performed with the R Bioconductor NOIseq package v2.14.0 (NOIseqBio) ([Tarazona et al., 2011](#)) and run with the R software v3.2.3 ([Team, 2015](#)). Because the RNAseq from the biofilm samples were derived from one pooled RNA preparation, the “no replicate parameter” was set ($pnr = 0.2$, $nss = 5$ and $\nu = 0.02$; pseudoreplicate generated) in NOIseq as described by [Tarazona et al. \(2011\)](#) under the NOIseq-sim section. Briefly, NOIseq-sim assumes (quoting) “that read counts follow a multinomial distribution, where probabilities for each gene in the multinomial distribution are the probability of a read to map to that gene”. Results from this statistical analysis showed that genes that had at least >2-fold higher transcript levels

from one type of cultures to the other showed significant differences. RNAseq reads from the planktonic pure cultures and the biofilm cultures were deposited in the SRA under the accession number [PRJNA525230](https://www.ncbi.nlm.nih.gov/sra/PRJNA525230). Annotations were based on services provided by GenBank (<https://www.ncbi.nlm.nih.gov/genbank>), RAST (Rapid Annotation using Subsystem Technology; <http://rast.nmpdr.org>) and KEGG (Kyoto Encyclopedia of Genes and Genomes; <https://www.genome.jp/kegg>) (Document S3).

To derive transcript reads not associated to *M. nitratireducenticrescens* and *H. nitrativorans*, reads were aligned to a concatenated sequence consisting of the three reference genomes (JAM1 + GP59 + NL23) and the two plasmids (from strain GP59). The reads that did not align were kept. Unaligned reads were *de novo* assembled at the National Center for Genome Analysis web site (<https://galaxy.ncgas-trinity.indiana.edu>) by Trinity v. 2.4.0 (Grabherr et al., 2011). These transcripts were deposited in SRA under the accession number [PRJNA525230](https://www.ncbi.nlm.nih.gov/sra/PRJNA525230). Estimation of the transcript abundance of the *de novo* assembled sequences was performed by RSEM (Li & Dewey, 2011). The resulting assembled sequences were annotated at the Joint Genomic Institute (<https://img.jgi.doe.gov/cgi-bin/m/main.cgi>) to find open reading frames with their putative function and affiliation (GOLD Analysis Project Id: Ga0307915, Ga0307877, Ga0307760). The annotations were then verified manually for discrepancies within the assembled sequences (Documents S4–S7).

RESULTS

Bacterial composition of the biofilm cultures by 16S rRNA gene sequencing

As reported by Payette et al. (2019), the original biofilm (OB) collected from the Biodome denitrification reactor was used as inoculum to colonize new carriers in a series of anoxic biofilm cultures cultivated under different conditions (Table 1; Fig. S1). We selected eight of these biofilm cultures for our present analysis for the following reasons. The Ref300N-23C biofilm cultures were used as reference for comparison analysis. The 300N-30C, 900N-23C and 900N-30C biofilm cultures had higher specific denitrification rates compared to the Ref300N-23C biofilm cultures. The 0%, 0.5% and 1.0% NaCl ASW biofilm cultures and the IO biofilm cultures were chosen because of the persistence of *H. nitrativorans* NL23 in these cultures as opposed to the other cultures. The composition of the bacterial community of these biofilm cultures and the OB was determined by sequencing the 16S rRNA genes to assess the impact of these specific conditions on the bacterial community (Fig. 1A; Table 2). The bacterial profiles of the OB and the IO biofilm cultures were distinct to each other, and to the seven biofilm cultures cultivated with different formulations in the ASW medium (Fig. 1B). The bacterial profiles of these latter cultures were however not very different because of the high proportions of *Methylophaga* spp. (>85%).

In the OB, high proportions of the 16S rRNA gene sequences were related to *Hyphomicrobium* spp. (45.8%) followed by *Oceanibaculum* spp. (12.3%), *Aquamicrobium* spp. (11.6%); *Methylophaga* spp. accounted for 3.5% (Table 2). In the IO biofilm cultures, the proportion of *Methylophaga* spp. was 12 times higher (42.8%) than in the OB, whereas it was 5.5 times lower for *Hyphomicrobium* spp. (8.4%). Higher proportions of *Marinicella*

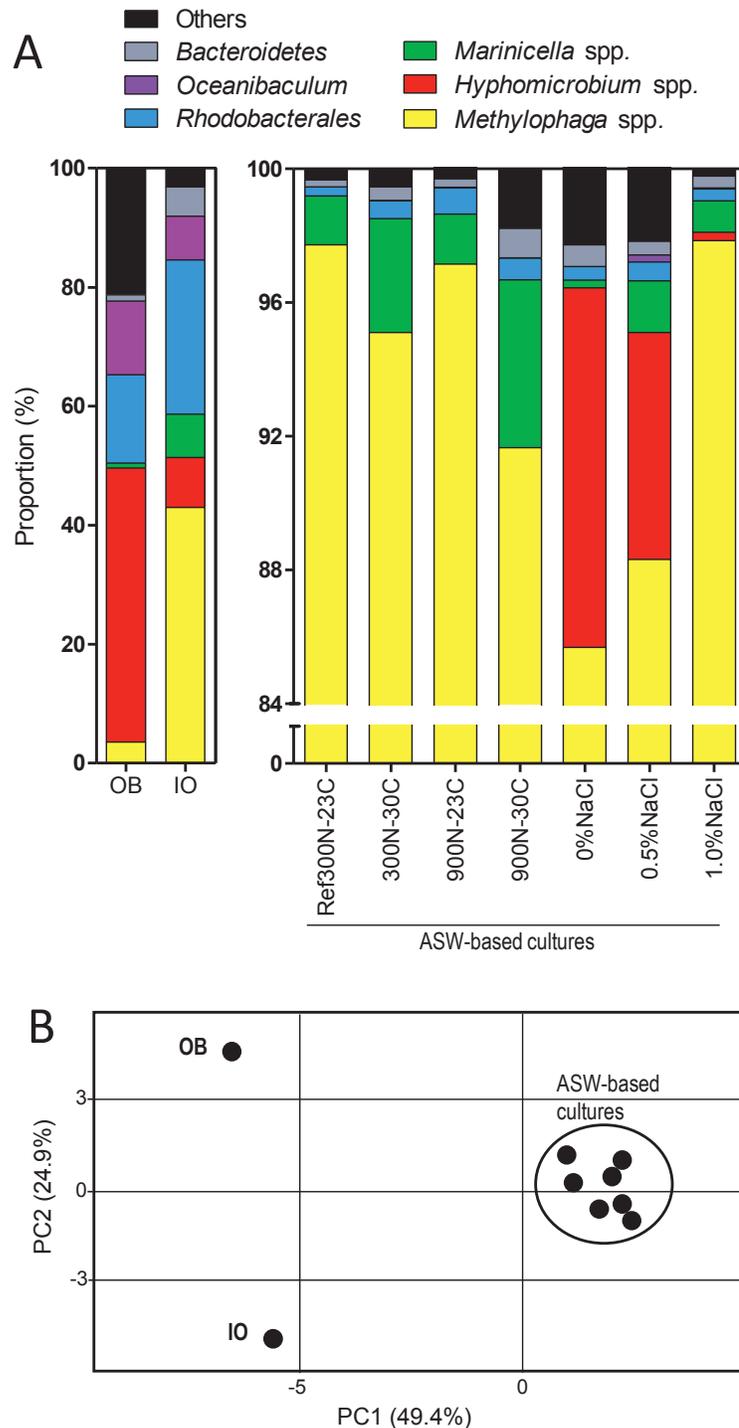


Figure 1 Proportion of affiliated OTUs in the biofilm cultures. (A) Bacterial composition of OB and the IO biofilm cultures was determined by sequencing the V6–V7–V8 variable regions of the 16S rRNA gene by Illumina, whereas the other samples were determined by sequencing the V1–V2–V3 variable regions by pyrosequencing. (B) Principal component analysis of the bacterial profiles of the biofilm cultures and the OB listed in Table 2.

Full-size DOI: [10.7717/peerj.7467/fig-1](https://doi.org/10.7717/peerj.7467/fig-1)

spp. (7.2%) and *Winogradskyella* spp. (4.3%) along with a much lower proportion of *Aquamicrobium* spp. (0.44%) were observed in the IO biofilm cultures compared to the OB (Table 2).

In the biofilm cultures cultivated with the four combinations of NO_3^- /methanol concentrations and temperatures in ASW medium containing 2.75% NaCl (Ref300N-23C, 300N-30C, 900N-23C, 900N-30C), *Methylophaga* spp. accounted for >90% of the 16S rRNA gene sequences followed by *Marinicella* spp. with proportions ranging from 1.5% to 5.0% (Fig. 1; Table 2). No sequences were found affiliated to *Hyphomicrobium* spp. under these conditions. In the biofilm cultures cultivated at low NaCl concentrations (0% NaCl, 0.5% NaCl, 1% NaCl), *Hyphomicrobium* spp. accounted for 11.8%, 6.8% and 0.25%, respectively of the 16S rRNA gene sequences (Fig. 1; Table 2). *Methylophaga* spp. was still the dominant genus with more than 85% of the 16S rRNA gene sequences; 16S rRNA gene sequences affiliated to *Marinicella* spp. were also found in significant proportions (Fig. 1; Table 2). Finally, 16S rRNA gene sequences affiliated to *Stappia* spp. were found in all biofilm cultures and in the OB.

The 16S rRNA gene sequences from the OB and the IO biofilm cultures that were derived by Illumina sequencing generated several thousands of reads affiliated to *Hyphomicrobium* spp. and *Methylophaga* spp. (Table 3). This tremendous amounts of sequences allowed assessing the presence of species other than *H. nitratorans* and *M. nitratreducentrescens* in these two biofilms. The phylogenetic analyses performed on these sequences allowed regrouping the OTUs in three clusters for *Hyphomicrobium* spp., and also three clusters for *Methylophaga* spp. (Figs. S2A and S2B). The vast majority (>90%) of the 16S rRNA gene sequences associated to these OTUs were affiliated to *H. nitratorans* or *M. nitratreducentrescens*, respectively, in the OB and the IO biofilm cultures (Clusters 1, Table 3). A small proportion of the OTUs (clusters 2 and 3) were affiliated to other *Hyphomicrobium* or *Methylophaga*, which suggests that other members of these genera were present in these biomasses in very low proportions.

Isolation of denitrifying bacterial isolates from the biofilm cultures

The biomass of the Ref300N-23C biofilm cultures was dispersed on different nutrient agar plates to isolate denitrifying bacteria other than *H. nitratorans* strain NL23 and *M. nitratreducentrescens* strain GP59. Isolates affiliated to the genera *Marinobacter*, *Pseudomonas*, *Paracoccus*, *Roseovarius*, *Thalassobius*, *Winogradskyella*, *Aequorivita* and *Exiguobacterium* (Table 4) were recovered from the Marine medium 2216 plates, which contains yeast extract and peptone (Atlas, 1993). Only isolates affiliated to the genera *Marinobacter* and *Paracoccus* showed consumption of NO_3^- and NO_2^- and production of gas, suggesting that they possess the complete denitrification pathway. The three isolates affiliated to the *Paracoccus* spp. have identical 16S rRNA sequences with the one of *Paracoccus* sp. strain NL8 that was isolated from the Biodome denitrification reactor (Labbé et al., 2003). This result suggests that strain NL8 persisted in the biofilm cultures. One representative of *Paracoccus* isolates (GP3) could grow with methanol as sole source of carbon; *Marinobacter* sp. GP2 could not.

Table 2 Most probable affiliation of 16S rRNA gene sequences in the biofilm cultures and the original biofilm.

Affiliation	Biofilm cultures (proportion %)										
	OB	IO	Ref300N-23C	900N-23C	300N-30C	900N-30C	0%NaCl	0.5%NaCl	1.0%NaCl		
<i>Ignavibacteria; Ignavibacteriales;</i>	<i>Ignavibacterium</i>	0.37	0.13		0.01	0.19	0.06				
<i>Tenericutes; Acholeplasmatales;</i>	<i>Acholeplasma</i>			0.05	0.04	0.05		0.04	0.52	0.02	
<i>Bacteroidetes; Flavobacteriales;</i>	<i>Aequorivita</i>	0.01	0.18		0.10	0.14	0.04	0.01	0.11	0.22	
" "	<i>Muricauda</i>	0.06	0.08			0.01	0.01		0.10		
" "	<i>Winogradskyella</i>	0.28	4.28	0.10	0.11	0.15	0.11		0.20	0.11	
<i>Proteobacteria;</i>											
<i>Alphaproteobacteria; Rhizobiales;</i>	<i>Aminobacter</i>	0.33	0.01		0.05				0.22		
" "	<i>Aquamicrobium</i> ^a	11.6	0.46					0.02			
" "	<i>Hoeflea</i>	0.30	0.76	0.03	0.01			0.06	0.12	0.06	
" "	<i>Nitratireductor</i>	1.40	0.03								
" "	<i>Hyphomicrobium</i> ^a	45.8	8.40					10.8	6.78	0.25	
	<i>H. nitrativorans</i> NL23, qPCR (cp <i>napA</i> /ng)	8.7(7.2) 10 ⁴	7.0(4.3) 10 ⁴	1.8(1.8) 10 ²	6.0(3.2) 10 ¹	1.3(0.5) 10 ²	1.3(0.9) 10 ²	2.8(1.0) 10 ⁴	5.3(6.5) 10 ⁴	1.1(0.9) 10 ⁴	
<i>Alphaproteobacteria; Rhodobacterales;</i>	<i>Litorisediminicola</i>	0.38	1.43								
" "	<i>Lutimaribacter</i>	1.71	8.93								
" "	<i>Marinovum</i>	1.28	1.62								
" "	<i>Maritimibacter</i>	2.50	2.44		0.01				0.01		
" "	<i>Oceanicola</i>	0.06	0.01			0.08	0.03		0.01		
" "	<i>Paracoccus</i> ^a			0.01	0.07	0.01			0.02		
" "	<i>Roseovarius</i>	7.32	8.18					0.01			
" "	<i>Stappia</i> ^a	0.02	1.26	0.09	0.43	0.27	0.42	0.27	0.35	0.17	
<i>Alphaproteobacteria; Rhodospirillales</i>	<i>Oceanibaculum</i>	12.3	7.36		0.03	0.01	0.01		0.20	0.04	
<i>Proteobacteria;</i>											
<i>Gammaproteobacteria; Alteromonadales</i>	<i>Marinobacter</i> ^a	0.15	0.05	0.06	0.05	0.02			0.01	0.01	
" "	<i>Oceanospirillales</i>	<i>Marinicella</i>	0.78	7.24	1.46	1.50	3.40	5.02	0.23	1.55	0.94
" "	<i>Pseudomonadales</i>	<i>Pseudomonas</i> ^a		0.09	0.11	0.04	0.13	0.52	0.64	0.80	0.05
" "	<i>Thiotrichales</i>	<i>Methylophaga</i>	3.50	42.8	97.7	97.1	95.1	91.8	85.7	88.3	97.9
	<i>M. nitratireducentescens</i> , qPCR (cp <i>narG1</i> /ng)	5.6(3.2) 10 ³	5.6(4.6) 10 ⁴	2.3(0.6) 10 ⁵	1.9(0.5) 10 ⁵	2.1(0.4) 10 ⁵	2.1(1.4) 10 ⁵	4.7(2.0) 10 ⁴	3.6(2.7) 10 ⁴	1.5(0.2) 10 ⁵	
Others		9.86	4.27	0.25	0.37	0.53	2.00	2.28	0.69	0.25	
Total of reads		348,358	319,265	9,610	14,048	8,532	13,915	13,800	9,375	12,560	

Notes.

The V1–V3 regions of the 16S rRNA gene sequences were amplified and sequenced by pyrosequencing except for the OB and IO samples from which the V6–V8 regions were sequenced by Illumina (see Document S2 for complete analysis and sequences).

OB, Original biofilm.

In grey: qPCR results are from Payette et al. (2019), with standard deviation values under parentheses of triplicate cultures. *narG1* used in qPCR for *M. nitratireducentescens* cannot discriminate strain JAM1 and strain GP59 (identical sequences between the two).

^aIdentified genus with species that were reported involved in denitrification.

Table 3 16S Operational Taxonomic Units (OTUs) affiliated to *Hyphomicrobium* spp. and *Methylophaga* spp. in the OB and the IO biofilm cultures.

		Number of OTUs	Number reads		Proportion of reads %	
			OB	IO	OB	IO
Sequence identity with <i>H. nitratorans</i>						
Cluster 1	95–100%	12	147,209	26,239	92.3	97.8
Cluster 2	91–97%	5	10,779	530	6.8	2.0
Cluster 3	89–92%	3	1,537	51	0.96	0.19
Sequence identity with <i>M. nitratreducentescens</i>						
Cluster 1	94–100%	18	11,809	136,197	96.4	99.7
Cluster 2	92.9–93.6%	3	321	284	2.62	0.21
Cluster 3	93.4–94.1%	3	115	65	0.94	0.05

Notes.

Cluster classification is based on phylogenetic analyses of the 16S rRNA gene sequences affiliated to *H. nitratorans* and *M. nitratreducentescens* (see [Tables S1](#) and [S2](#), and [Figs. S2A](#) and [S2B](#)).

Table 4 Affiliation of the isolates isolated from the reference biofilm cultures (Ref300N-23C).

Isolates	Denitrifying activities ^a
<i>Alphaproteobacteria, Rhodobacterales</i>	
<i>Rhodobacteraceae</i> GP11	No
<i>Paracoccus</i> sp. GP3, GP8, GP20	Full
<i>Roseovarius</i> sp. GP9, GP10, GP13, GP14	No
<i>Roseovarius</i> sp. GP12	No
<i>Thalassobius</i> sp. GP19	No
<i>Gammaproteobacteria</i>	
<i>Marinobacter</i> sp. GP1, GP2, GP24	Full
<i>Pseudomonas</i> GP41	Nitrate
<i>Bacteroidetes, Flavobacteriales</i>	
<i>Aequorivita</i> sp. GP15	No
<i>Winogradskyella</i> sp. GP16, GP18	No
<i>Bacillales</i>	
<i>Exiguobacterium</i> sp. GP46	No

Notes.

^aFull: nitrate and nitrite are consumed. Nitrate: only nitrate was consumed.

Metatranscriptomic analysis of the biofilm cultures

The metatranscriptomic approach has allowed assessing the contributions of the microbial community to the metabolic processes in the biofilm cultures. We have chosen to focus on three biofilm cultures, which were the Ref300N-23C (the reference biofilm cultures), 900N-30C (highest denitrification rates; [Table 1](#)) and 0% NaCl (persistence of *H. nitratorans* strain NL23) biofilm cultures. Because the genomes of *H. nitratorans* strain NL23 and *M. nitratreducentescens* strain JAM1 and strain GP59 were available, we first determined changes in the transcript levels of genes associated to these genomes between the biofilm cultures and the planktonic pure cultures of the respective strains. To assess the contribution of other microorganisms in biofilm cultures, the reads from the metatranscriptomes that

did not align with the three reference genomes were used to derive *de novo* assembled transcripts. These transcripts were annotated for function and bacterial affiliation.

Gene expression profiles of *M. nitratireducenticrescens* strain GP59 in the biofilm cultures

Because >80% of the genomes of strains JAM1 and GP59 are identical, high proportions of reads from the biofilm metatranscriptomes can align to both genomes. [Geoffroy et al. \(2018\)](#) showed that the gene expression profiles of the common genes between both strains in planktonic pure cultures were similar. In [Payette et al. \(2019\)](#), the concentrations of strain GP59 and strain JAM1 in the biofilm cultures were determined by qPCR. In the three selected biofilm cultures for the metatranscriptomic analysis, the concentrations of strain GP59 (copies of *nirK* by ng biofilm DNA) were one to three orders of magnitude higher than those of strain JAM1 (copies of *tagH* by ng biofilm DNA). Because of these differences, it was assumed that most of the transcript reads associated to *M. nitratireducenticrescens* in the biofilm cultures were from strain GP59. Metatranscriptomic analysis in relation with strain JAM1 is described in [Documents S8](#) and [S11](#). The transcriptome of strain GP59 was also derived from planktonic pure cultures cultivated under anoxic conditions in the *Methylophaga* 1403 medium ([Fig. S1](#)). The choice of this medium was because suboptimal growth occurred with strain GP59 in ASW medium. The relative transcript levels of the corresponding genes in the biofilm cultures and the planktonic pure cultures were compared to assess changes in the metabolisms of the strain that occurred between the two environments. All quantitative changes described below of the transcript levels in the biofilm cultures are expressed relative to the transcript levels in the planktonic pure cultures.

Among all genes of strain GP59, between 11% and 21% of them had higher relative transcript levels in the biofilm cultures. At the opposite, 6 to 17% of all genes of strain GP59 were expressed at higher relative transcript levels in planktonic pure cultures ([Fig. 2](#)). Strain GP59 contains two plasmids, and most of the genes encoded by these plasmids had much lower relative transcript levels in the biofilm cultures ([Fig. 2](#)). Genes involved in the nitrogen metabolism and iron transport were globally at higher relative transcript levels in the biofilm cultures ([Table 5](#); [Figs. 2](#) and [3](#)).

For the denitrification genes, *narXL* encoding the regulatory factors of the *nar* systems showed no differences between the biofilm cultures and the planktonic pure cultures in the relative transcript levels ([Table 5](#)). Small upregulation of the *nar2* operon with about 3-fold increases in relative transcript levels occurred in the biofilm cultures. These levels were lower in the 900N-30C biofilm cultures for the *nar1* operon and were about the same levels in the two other biofilm cultures and the planktonic pure cultures. The *nor1* operon had the same relative transcript levels in the 300N-23C and 900N-30C biofilm cultures and the planktonic pure cultures, and a 3-fold decrease was noticed in these levels in the 0% NaCl biofilm cultures ([Table 5](#)). No significant changes in the expression of the *nos* operon occurred between both types of cultures. The relative transcript levels of *nirK* were 5 to 10-times higher in the 300N-23C and 900N-30C biofilm cultures, whereas these levels were similar in the 0% NaCl biofilm cultures and the planktonic pure cultures ([Table 5](#)). Higher

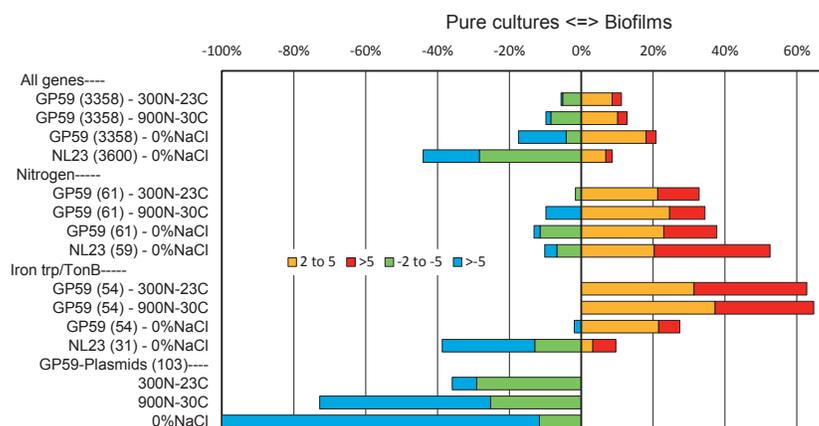


Figure 2 Relative expression profiles of *M. nitratireducens* GP59 and *H. nitrativorans* NL23 in biofilm cultures. All the deduced amino acid sequences associated to the genome and plasmids of strain GP59 and the genome of strain NL23 were submitted to the BlastKOALA (genome annotation and KEGG mapping) at the Kyoto encyclopedia of genes and genomes (KEGG). Genes associated to specific metabolisms were sorted out and the corresponding ratios of the Biofilm Transcripts Per Million (TPM) versus the pure culture TPM were derived. When the ratios were < 1 , the negative inverse values ($-1/\text{ratio}$) were calculated. Data are expressed as the percentage of genes in each category that are more expressed in the biofilm cultures (right, 2–5 times, and > 5 times) or in pure cultures (left, -2 to -5 times and > -5 times). Number within parentheses is the number of genes involved in the selected pathways. Other metabolic profiles are detailed in Figs. S3 and S4.

Full-size DOI: 10.7717/peerj.7467/fig-2

relative transcript levels of genes involved in the ammonium transport and the assimilatory $\text{NO}_3^-/\text{NO}_2^-$ reduction pathway were observed in the biofilm cultures (Table 5; Fig. 3). Absence of nitrogen source other than NO_3^- in the ASW medium and presence of 37 mM NH_4^+ in the medium used for the planktonic pure cultures (*Methylophaga* 1403) could explain these differences in the assimilatory pathway.

Figure 3 illustrates changes in the relative expression profiles in the biofilm cultures of major pathways in strain GP59. The relative transcript levels of *mxoFJGI* encoding the small and large subunits of the methanol dehydrogenase (MDH) and the cytochrome c-L increased by 2–6 folds in biofilm cultures. Two out of the four *mxoF*-related products (*xoxF*) showed 2- to 9-fold decreases in their relative transcript levels in the biofilm cultures. As observed in *Methylorubrum extorquens*, the genome of *M. nitratireducens* strain GP59 encodes three formate dehydrogenase with the same gene arrangement (Chistoserdova et al., 2004). The *fdhCBAD* operon that encodes the NAD-linked, Mo-formate dehydrogenase showed ca. 60-fold increases in the relative transcript levels in the biofilm cultures, whereas the two other *fdh* operons stayed at the same levels of those of the planktonic pure cultures. Genes encoding NAD-dependent formate dehydrogenase were upregulated in biofilms formed by *Desulfovibrio vulgaris* compared to planktonic cultures (Clark et al., 2012). This was also the case in biofilms formed by *Staphylococcus aureus* where the NAD-dependent formate dehydrogenase genes were among the highest upregulated genes (Resch et al., 2005). In both studies, this upregulation correlated with increase in formate dehydrogenase activity. Contrary to planktonic cultures, accumulation of formate

Table 5 Changes in the relative transcript levels of genes involved in the nitrogen metabolism in strain GP59 and strain NL23.

Genes	Ratio TPM Biofilm cultures/TPM pure cultures				
	GP59			NL23	
	Ref300N-23C	900N-30C	0%NaCl	0%NaCl	Genes
Denitrification					
<i>narXL</i>	ns	ns	ns	−9.4	<i>napGH</i>
<i>narK1K2GHJI-1</i>	ns	−7.4	2.1	8.2	<i>napEFDABC</i>
<i>narK12</i>	ns	ns	2.0		
<i>narGHJI-2</i>	2.7	2.8	3.0		
<i>nirK</i>	11	4.7	ns	49	<i>nirKV</i>
<i>norCBDQ-1</i>	ns	ns	−2.7	5.9	<i>norCBQDE</i>
<i>norRE</i>	ns	ns	−3.1		
<i>norCBQD-2</i>	2.4	3.8	2.8		
<i>nosRZDFYL</i>	ns	ns	ns	5.2	<i>nosRZDFYLX</i>
<i>nnrS</i> (3)	ns	ns	ns	ns	<i>nnrS</i> (2)
<i>nsrR</i> (2)	2.1	2.5	ns	ns	<i>nsrR</i>
<i>DnrN</i>	2.3	2.8	−3.0	ns	<i>nnrU</i>
NOdiox (2)	ns	ns	ns	4.0	<i>nnrR</i>
Nitrogen assimilatory pathway					
				ns	<i>nasTS</i>
NO ₃ /NO ₂ trp	6.0	7.2	17.9	7.8	<i>ntrABC</i>
<i>nasAnirBD</i>	12	24	42	3.3	<i>nirBANasA</i>
<i>ntrYX</i>	ns	ns	ns	ns	<i>ntrYX</i>
<i>glnA</i>	2.0	3.1	5.1	19	<i>glnA</i>
				−2.1	<i>glnA</i> (3)
<i>gltBD</i> : GOGAT	ns	ns	ns	ns	<i>gltBD</i>
<i>glnB</i>	ns	ns	ns	14	<i>glnB</i>
GDH	ns	ns	ns	ns	GDH
<i>glnD</i>	ns	ns	ns	−2.9	<i>glnD</i>
<i>glnE</i>	ns	ns	ns	−3.2	<i>glnE</i>
<i>glnK</i>	18	24	25	22	<i>glnK</i>
NH ₄ trp	11	11	11	20	NH ₄ -trp
<i>glnLG</i>	ns	2.1	5.9	ns	<i>glnLG</i>

Notes.

Data values are the ratios of the biofilm-culture TPM divided by the planktonic pure-culture TPM. When the ratios were <1, the negative inverse values (−1/ratio) were calculated. Positive ratios mean higher relative transcript levels in the biofilm cultures, and negative ratios higher relative transcript levels in the planktonic pure cultures. ns, no significant changes; *glnA*, glutamine synthetase; *gltBD*, glutamate synthase; GDH, glutamate dehydrogenase; *glnB*, nitrogen regulatory protein P-II 1; *glnK*, nitrogen regulatory protein P-II 2; *glnD*, uridylyltransferase; *glnLG*, nitrogen regulation sensor histidine kinase and response regulator; trp, transporter; Diox, dioxygenase; *nnrS*, involved in response to NO; *nsrR*, NO-sensitive transcriptional repressor; *DnrN*, NO-dependent regulator; *nasAnirBD*, assimilatory nitrate and nitrite reductase; *ntrXY*, Nitrogen regulation proteins; *narXL*, nitrate/nitrite sensor histidine kinase and response regulator; *narK*, nitrate/nitrite transporter; *nosRE*, NO-reductase transcription regulator and activation protein.

could occur in cell vicinity in the biofilm that would be toxic for the cells (Resch et al., 2005). Therefore, upregulation of *fdhCBAD* operon could be related to detoxification. The relative transcript levels of the gene encoding the cytochrome c555 were 40–70 times higher in the biofilm cultures. Genes encoding the other cytochromes, pseudoazurins and azurin were

expressed at similar levels in both types of cultures. Genes involved in the formaldehyde metabolism to formate and CO₂, glycolysis, the ribulose monophosphate pathway, the Entner Doudorof pathway, the tricarboxylic acid cycle, and the pentose pathway showed their relative transcript levels in general unchanged. Few genes in these pathways had 2- to 11-fold differences between the planktonic pure cultures and the biofilm cultures. The genome of strain GP59 encodes the major enzymes involved in the Calvin-Benson-Bassham cycle: the ribulose-bisphosphate carboxylase (Rubisco) and the phosphoribulokinase (Prk). In the 0% NaCl biofilm cultures, the relative transcript levels of the Rubisco gene operon (*rbcSL*) jumped by 66 times. This upregulation was less pronounced in the Ref300N-23C biofilm cultures (3-fold increase). For the *prk* gene, the relative transcript levels were 3 to 4 times higher in the 0% NaCl biofilm cultures. The nature of this upregulation is unknown. Except for the cytochrome *c555*, the relative transcript levels of genes encoding for the oxidative phosphorylation metabolism were unchanged. Several genes involved in iron transport showed higher relative transcript levels in the biofilm cultures (2- to >50-fold increases). The nature of this upregulation in the biofilm cultures is unknown, as the biofilm and planktonic pure cultures were cultivated with trace elements containing iron (Payette *et al.*, 2019).

Gene expression profiles of *H. nitrativorans* strain NL23 in the 0% NaCl biofilm cultures

As with *M. nitratireducenticrescens* strain GP59, the transcript levels of genes associated to *H. nitrativorans* strain NL23 were compared between the biofilm cultures and the planktonic pure cultures to assess changes in the metabolisms that occurred between the two environments. The planktonic pure cultures of strain NL23 were cultivated under anoxic conditions in the 337a medium (Fig. S1). The choice of this medium was because strain NL23 could not grow in ASW (with 2.75% NaCl).

The overall analysis of the three metatranscriptomes confirmed the results obtained by qPCR assays (Payette *et al.*, 2019) and the 16S rRNA gene analysis (Table 2). High number of reads (40×10^6) derived from the metatranscriptome of the 0% NaCl biofilm cultures aligned with the NL23 genome, but <20,000 reads derived from the Ref300N-23C and 900N-30C metatranscriptomes did. In the 0% NaCl biofilm cultures, <10% of all NL23 genes had a higher relative transcript levels in the 0% NaCl biofilm cultures, whereas this was the case for >40% genes in planktonic pure cultures (Fig. 2). These results suggest important changes had occurred in the regulation of gene expression between the planktonic pure cultures and the biofilm cultures. Genes involved in the energy (Fig. S4) and nitrogen metabolisms (Fig. 2; Table 5) had globally higher relative transcript levels in the 0% NaCl biofilm cultures (Fig. 2).

Higher relative transcript levels (5- to 8-times) for the *nap*, *nor* and *nos* operons were observed in the 0% NaCl biofilm cultures (Table 5). *nirK* was highly upregulated in the biofilm cultures with 49-fold increase in the relative transcript levels (Table 5). The *napGH* operon however had a 9.4-fold decrease in the relative transcript levels in the 0% NaCl biofilm cultures. As observed with strain GP59, substantial changes in the relative transcript levels of genes involved in the ammonium transport and the assimilatory NO₃⁻/NO₂⁻

reductase were observed with 3- to 22-fold increases in these levels (Table 5, Fig. 4). These results correlate with the absence of NH_4^+ in the ASW medium, and thus NO_3^- – the only source of N, compared to the 337a medium used for the planktonic pure cultures, which contains 3.8 mM NH_4^+ .

Figure 4 illustrates changes in relative expression profiles of major pathways in the 0% NaCl biofilm cultures of strain NL23. The relative transcript levels of *mxoFJGI* increased by 2 folds in the biofilm cultures. The relative transcript levels of the *mau* operon (methylamine dehydrogenase) showed a 15-fold decrease in the biofilm cultures. The nature of such decrease is unknown as strain NL23 was not fed with methylamine in any of our cultures. The three *xoxF* genes did not show substantial changes in their transcript levels in both types of cultures. Genes involved in the formaldehyde metabolism to formate and CO_2 , glycolysis, the tricarboxylic acid cycle, and the pentose pathway showed their transcript levels in general unchanged between the planktonic pure cultures and the biofilm cultures. Few genes in these pathways had 2- to 5-fold differences in their relative transcript levels. The two genes encoding the key enzymes in the serine pathway (alanine-glyoxylate transaminase, glycine hydroxymethyltransferase) had 3- to 7-fold increases in their relative transcript levels in the biofilm cultures. As *Methylorubrum extorquens*, the NL23 genome encodes the ethylmalonyl-CoA pathway (Chistoserdova et al., 2003; Peyraud et al., 2009), which did not show changes overall in the transcript levels of the corresponding genes between the two types of cultures. Contrary to *M. extorquens* however, a gene encoding the isocitrate lyase is present in strain NL23 and showed a 25-fold upregulation in the biofilm cultures. The isocitrate lyase is one of the key enzymes of the glyoxylate bypass that catalyzes the transformation of isocitrate to succinate and glyoxylate. Gene encoding isocitrate lyase is also present in other available *Hyphomicrobium* genomes. All these results suggest that in the 0% NaCl biofilm cultures, the carbon metabolism increased in activity and that the glycine regeneration for the serine pathway by the glyoxylate was upregulated. Among genes involved in the oxidative phosphorylation, the relative transcript levels were higher (2–13 times) in the biofilm cultures with those encoding the NADH dehydrogenase, the cytochrome c reductase, with one of the cytochromes c and the F-type ATPase. Combined with increases in the relative transcript levels of the denitrification and the carbon pathways, these results suggest that increases in electron donor activities correlates with the need of electron for the nitrogen dissimilatory metabolism in the biofilm cultures. Strain NL23 possesses four types of cytochrome oxidase (aa3, bo, bd-I and cbb3) (reduction of O_2 in H_2O) that were about expressed at the same levels in both types of cultures. Figure 4 also illustrates the dynamic changes of transporters and two component systems. Several of these transporters had lower relative transcript levels in the 0% NaCl biofilm cultures. Contrary to strain GP59, genes involved in iron transport were not strongly affected in their gene expression in the 0% NaCl biofilm cultures (Figs. 2 and 4).

The composition of the active microbial community in the biofilm cultures

As mentioned above, the reads from the three metatranscriptomes that did not align with the genomes of *H. nitrativorans* strain NL23 and *M. nitratireducenticrescens* strain

GP59 and strain JAM1 were *de novo* assembled. These reads were subsequently aligned to the *de novo* assembled transcripts to derive the relative levels of these transcripts in the biofilm cultures. The *de novo* assembled sequences were then annotated for function and affiliation. Finally, these sequences were grouped by microbial affiliation to determine the active populations in the biofilm cultures and to assess their level of involvement in these biofilm cultures (Table 6).

It was estimated that between 5 and 10% reads of the three metatranscriptomes were derived from other microorganisms than *H. nitratorans* strain NL23 and *M. nitratreducenticrescens* strain GP59 and strain JAM1 (Document S4). The proportions of transcripts affiliated to Archaea and Eukarya accounted together for <0.1% (Table 6), which suggests very low abundance of these microorganisms in the biofilm cultures. The proportions of transcripts affiliated to viruses, phages and plasmids in the *de novo* assembled transcripts represented between 0.6 and 21.7% (Table 6).

Twenty-seven bacterial taxa were selected for their overall transcript levels in at least one of the three biofilm cultures (Table 6). All the taxa detected by the 16S rRNA gene sequencing are present in this list (Table 2 and Document S2). These 27 taxa represented between 22% and 35% of the *de novo* assembled transcripts. Among these taxa, genes encoding the four denitrification reductases were present in the *de novo* transcripts affiliated to *Marinobacter* spp., *Stappia* spp. and *Pseudomonas* spp. However, only *de novo* assembled transcripts affiliated to *Stappia* spp. showed the complete set of denitrification genes in the three biofilm cultures and organized in operons (*napABC*, *napADFE*, *norCBQD*, *nosRZDF*).

Further analysis of the expression profiles of the 27 bacterial taxa was performed to assess whether some taxa were influenced in their global metabolic activities by the specific conditions of the biofilm cultures. The overall transcript levels of the 27 bacterial taxa (Table 6) were compared between each biofilm culture by clustering analysis (Fig. 5). NaCl concentration was the main factor of clustering as two distinct clusters were derived. The low salt cluster consisting of nine bacterial taxa showed higher relative transcript levels in the 0% NaCl biofilm cultures, whereas the marine cluster of 13 bacterial taxa had higher relative transcript levels in the Ref300N-23C and 900-30C biofilm cultures. A third cluster showed five bacterial taxa with lower relative transcript levels in the 900N-30C biofilm cultures compared to the 0% NaCl and Ref300N-23C biofilm cultures. In these cases, higher temperature (30 °C vs 23 °C) and higher NO₃⁻ and methanol concentration (64.3 mM vs 21.4 mM NO₃⁻; 0.45% vs 0.15% methanol) may have negatively affected these populations.

DISCUSSION

In the environment, numerous bacteria belonging to different taxa can accomplish denitrifying activities, and many of them were encountered in different types of denitrification processes (Lu, Chandran & Stensel, 2014). Very few studies describing the microbial community of methanol-fed denitrification systems have been reported so far. Most of these studies are based on cloned 16S rRNA gene sequences of around 100

Table 6 Microbial diversity and the associated denitrification genes in the biofilm cultures from the *de novo* assembled transcripts.

Affiliation	Biofilm cultures (TPM)			Denitrification genes
	Ref300N-23C	900N-30C	0%NaCl	
• Actinobacteria				
<i>Streptomyces</i>	6,103	729	2,499	–
• Bacteroidetes				
<i>Sunxiuqinia</i>	212	504	2,639	<i>norB</i>
<i>Geofilum</i>	77	109	1,1870	–
<i>Aequorivita</i>	18,218	25,569	1,398	<i>nirK, norB;C;D;Q, nosZ;DFY</i>
<i>Winogradskyella</i>	8182	7,917	169	<i>nirK, norC, nosZ;L;D</i>
<i>Lentimicrobium</i>	3032	3,370	1,671	–
<i>Xanthomarina</i>	190	4,555	9,598	<i>nirK, norB;D;C;Q, nosZ;L;D</i>
• Ignavibacteriales	5,513	1,4206	171	<i>napC;H</i>
• Tenericutes	6,367	5,003	13,409	–
• Alphaproteobacteria				
<i>Aminobacter</i>	18	438	5,385	<i>napA</i>
<i>Aquamicrobium</i>	23	456	22,835	<i>napA</i>
<i>Hoeflea</i>	3,882	87	1,365	<i>narG;H;J;I, nosZ;R;D, nirK</i>
<i>Hyphomicrobium</i>	0	1,679	3,0057	<i>narH;I, norB;C;Q;D;E, nosZ;D;R;F</i>
<i>Mesorhizobium</i>	411	1,050	12,505	<i>napA;D;E, nirK, nosZ</i>
<i>Paracoccus</i>	1,427	1,219	1,204	<i>nirS, narC;D;Q, nosR</i>
<i>Roseovarius</i>	1,2367	16,479	686	<i>nirS, nirK, norB;C;D;Q;E, nosZ;D;R</i>
<i>Stappia</i>	39,926	30,885	39,745	<i>napABC, napADFE-nnrS, nirK, norCBQD, nosRZDF;E</i>
<i>Maritimibacter</i>	6,363	8,360	10,000	<i>narI</i>
<i>Oceanibaculum</i>	18,115	9,071	25,544	<i>narG;H;J;I</i>
• Betaproteobacteria				
<i>Azoarcus</i>	48	120	2,854	<i>napA, nosZ</i>
• Deltaproteobacteria				
<i>Bradymonas</i>	73	9,652	11,178	–
• Gammaproteobacteria				
<i>Marinicella</i>	36,554	125,466	149,24	<i>nirS, norB;C;Q;D</i>
<i>Marinobacter</i>	14,545	1,122	264	<i>narG;H;J;I, norB;C;Q, nirK, nirS, nosZ;F;D;R</i>
<i>Methylophaga</i>	64,802	78,566	80,482	<i>narG;H;J;I, norB;C;D;Q;E, nosD;R</i>
<i>Idiomarina</i>	5,147	1,027	513	<i>narG;H;J, nirK, nirS, norB</i>
<i>Pseudomonas</i>	6,175	1,333	18,198	<i>narG;H;J;I, nirK, nirS, norB;C;D;Q, nosZ;R,</i>
<i>Wenzhouxiangella</i>	132	2,857	83	–
• Other bacteria	113,200	243,523	155,554	^a
• Archaea	113	297	303	
• Eukarya	181	352	416	
• Phages, viruses, plasmids	99,623	5,740	217,095	

(continued on next page)

Table 6 (continued)

Affiliation	Biofilm cultures (TPM)			Denitrification genes
	Ref300N-23C	900N-30C	0%NaCl	
• Unclassified	281,015	207,759	157,515	
• Transcripts with no genes	248,782	196,062	148,792	

Notes.

Reads that did not align to the three reference genomes and plasmids were *de novo* assembled. These reads were then aligned to these assembled sequences. The relative transcript levels of the assembled sequences in a metatranscriptome were expressed as transcripts per million (TPM). Putative genes from the assembled sequences were annotated for function and affiliation. The TPM of the genes affiliated to specific bacterial taxa were then summed. Denitrification genes identified by annotations from respective affiliated bacterial taxa were sorted out.

^aDenitrification genes were found scattered in other bacterial taxa.

clones or based on fluorescence *in situ* hybridization (Baytshtok *et al.*, 2008; Ginige *et al.*, 2004; Hallin *et al.*, 2006; Neef *et al.*, 1996; Osaka *et al.*, 2008; Osaka *et al.*, 2006; Rissanen *et al.*, 2016; Rissanen *et al.*, 2017; Sun *et al.*, 2016; Yoshie *et al.*, 2006). In all these studies, high proportions of *Hyphomicrobium* spp. were found in combination with high proportions of other methylotrophs such as *Methyloversatilis* spp., *Methylophilus* spp., *Methylotenera* spp. or *Paracoccus* spp. The Biodome marine denitrification system showed no exception to this trend with co-occurrence of *Hyphomicrobium* spp. and the marine methylotroph *Methylophaga* spp. This co-occurrence was also observed in two other denitrification systems treating saline effluents (Osaka *et al.*, 2006; Rissanen *et al.*, 2016) (see ‘Discussion’ by Payette *et al.* (2019)). The bacterial diversity of the biofilm taken from the Biodome denitrification system was assessed before when the reactor was operational in 2002 by deriving a 16S rRNA gene library and by culture approach (Labbé *et al.*, 2003; Labbé, Parent & Villemur, 2004). Beside *Hyphomicrobium* sp. and *Methylophaga* sp., *Paracoccus* sp., *Sulfitobacter* sp., *Nitratireductor aquibiodomus*, and *Delftia* sp. among others were identified. In the present report, a more complete determination of the composition of the bacterial community of the denitrifying biofilm that was frozen in 2006 (when the denitrification system was dismantled by the Biodome) was possible with the new sequencing technology.

The composition of the bacterial community of the OB and the IO biofilm cultures was derived from a different region of the 16S rRNA genes (V6–V8) than that used for the other biofilm cultures (V1–V3), and each region was sequenced by a different technology (Illumina and pyrosequencing, respectively). Pyrosequencing technology was no longer available at the time of the sampling of the OB and the IO biofilm cultures, which were carried two years later than the other biofilm cultures. Despite these differences, we believe that the results generated by these two approaches were comparable because they are consistent with the results obtained by qPCR assays (Table 2) that have determined the concentrations of *M. nitratireducenticrescens* (copies *narG1* per ng biofilm DNA) and *H. nitrativorans* strain NL23 (copies *napA* per ng biofilm DNA) in the biofilm cultures (Payette *et al.*, 2019). For instance, qPCR showed very low levels of *H. nitrativorans* strain NL23 in the Ref300C-23C biofilm cultures and high level in the OB. These results concur with the absence of 16S rRNA sequences associated to *Hyphomicrobium* spp. in the Ref300C-23C biofilm cultures (pyrosequencing) and high number of these sequences in the OB (Illumina sequencing).

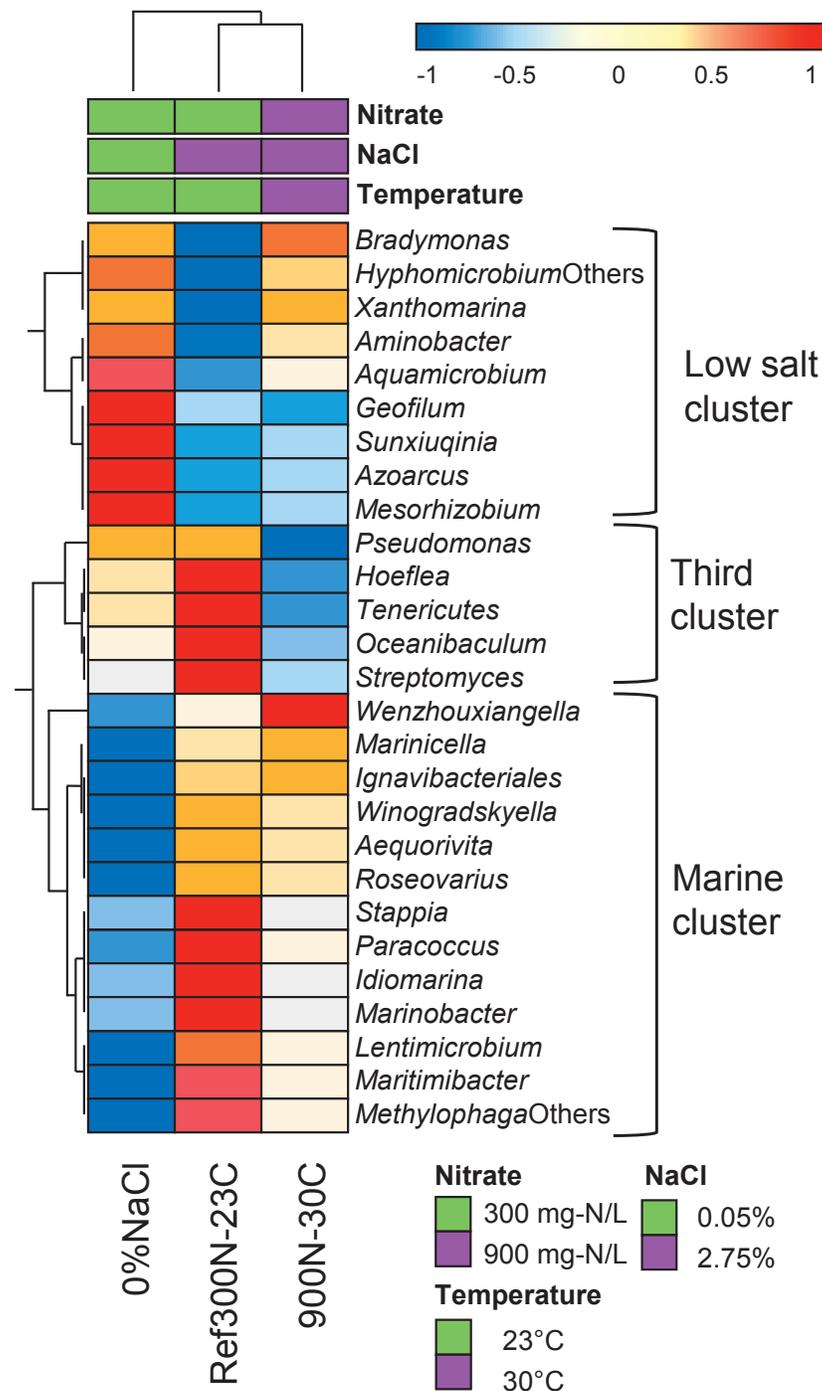


Figure 5 Hierarchical clustering of selected bacterial taxa in biofilm culture metatranscriptomes. Heatmap represents differences in the overall gene expression patterns (expressed as TPM; from Table 6) (\log_{10} [TPM by geometric average of TPM]) between the three biofilm cultures for the respective bacterial taxa. Analysis was performed at ClustVis web site (<https://biit.cs.ut.ee/clustvis/>) (Metsalu & Vilo, 2015).

Full-size DOI: 10.7717/peerj.7467/fig-5

The OB cultivated under the different conditions in the ASW medium showed important changes in the *Methylophaga* and the *Hyphomicrobium* populations. The proportion of 16S rRNA gene sequences associated to *Methylophaga* spp. was 3.5% in the OB but was very high, between 85% and 97%, in these cultures. On the contrary, the proportion of 16S rRNA gene sequences associated to *Hyphomicrobium* spp. was high (46%) in the OB, but was very low (0% to 11%) in these cultures. The NaCl concentrations in the ASW had an impact on the *Hyphomicrobium* populations. 16S rRNA gene sequences associated to *Hyphomicrobium* spp. were absent in the biofilm cultures cultivated in ASW at 2.75% NaCl (Ref300N-23C, 300N-30C, 900N-23C, 900N-30C), but were present in the biofilm cultures cultivated at low NaCl concentrations (0%, 0.5% and 1%). These results concur with those obtained by qPCR for the concentrations of *M. nitratireducenticrescens* and *H. nitrativorans* strain NL23 (Payette et al., 2019). Cultivating the OB in the IO medium had a different impact on the *Hyphomicrobium* populations. Although the concentrations of salts in the IO medium and in the ASW with 2.75% NaCl were similar (around 3.5%), persistence of *Hyphomicrobium* spp. occurred in the IO biofilm cultures. In fact, the concentrations of *H. nitrativorans* strain NL23 determined by qPCR between the OB and the IO biofilm cultures were at the same levels of magnitude. The lower proportion of 16S rRNA gene sequences associated to *Hyphomicrobium* spp. in the IO biofilm cultures compared to OB was a consequence of the substantial growth of *M. nitratireducenticrescens* in these cultures, with a 10-fold increase in concentration as revealed by qPCR (Table 2) (see Payette et al., 2019 for further discussion). Grob et al. (2015) also observed strong growth of *Methylophaga* spp. in their seawater samples that were fed with 100 μ M methanol with relative proportions of 16S rRNA gene sequences raising from <0.5% at $T = 0$ to 84% after 3 days.

Cultivating the OB in higher concentrations of NO_3^- and methanol (64.3 mM/0.45%; C/N 1.5) and/or at 30 °C (300N-30C, 900N-23C and 900N-30C biofilm cultures) resulted in increases of 20% to 85% in the specific denitrification rates compared to the Ref300N-23C biofilm cultures (Table 1) (Payette et al., 2019). Temperature was shown to be the main factor of these increases. However, raising the NO_3^- and methanol concentrations or/and temperature in these cultures did not have an important impact on the bacterial community when compared to the Ref300N-23C biofilm cultures. Metatranscriptomic analysis of the Ref300N-23C and 900-30C biofilm cultures did not reveal either substantial changes in the gene expression profiles between these two cultures. The higher specific denitrification rates of these biofilm cultures could be related to higher metabolisms at the protein levels such as the processing of NO_3^- by the reductases and transporters.

Results from the 16S rRNA gene sequences showed that *Marinicella* spp. were present in the OB and all the biofilm cultures, and were the second most abundant bacterial population in the biofilm cultures cultivated in ASW at 2.75% NaCl. These results concur with the metatranscriptomes of the biofilm cultures where *Marinicella* spp. had the relative transcript levels among the highest in the *de novo* assembled transcripts. *Marinicella* spp. are considered strict aerobic bacteria with no indication of NO_3^- reduction (Romanenko et al., 2010) although a previous study reported high relative abundances of *Marinicella* spp. in anoxic sulfide oxidizing reactors in which nitrate was used as the electron acceptor

(Huang et al., 2015). Genome annotations of two *Marinicella* strains (GenBank: *Marinicella* sp. F2—assembly number ASM200005v1 and *M. litoralis* KMM 3900—ASM259191v1) (Wang et al., 2018) did not reveal complete denitrification pathway, beside a nitric oxide reductase gene cluster also detected here in our metatranscriptomes. Together with the presence of *nirS* gene (Table 6), these results indicated that *Marinicella* spp. might have the capacity to use intermediates of the denitrification cycle to support their growth.

The 16S rRNA gene sequences provided evidence of the presence of *Pseudomonas* spp., *Marinobacter* spp., *Stappia* spp., *Paracoccus* spp. and *Aquamicrobium* spp. in the OB and in the biofilm cultures. Some species belonging to these genera were reported to carry denitrification. Isolates affiliated to the genera *Pseudomonas*, *Marinobacter* and *Paracoccus* were recovered from the Ref300N-23C biofilm cultures. The *Marinobacter* and *Paracoccus* isolates could perform denitrifying activities and grow under anoxic conditions, whereas the *Pseudomonas* isolate could only consume NO_3^- .

Although denitrification genes were found in several of the bacterial populations identified by the metatranscriptomic approach, only transcripts encoding the four denitrification reductases affiliated to *Stappia* spp. were found in the three examined biofilm cultures. The proportions of *Stappia* spp. in the 16S rRNA gene sequences of these biofilm cultures ranged between 0.09% and 0.42% (Table 2). *Stappia* spp. are chemoorganotrophic bacteria found in marine environments (Weber & King, 2007) that can oxidize carbon monoxide. They possess the form I *coxL* gene encoding the large subunit of carbon monoxide (CO) dehydrogenase. Some also contain a gene for the large subunit of ribulose-1,5-bisphosphate carboxylase (RuBisCO) and may be able to couple CO utilization to CO_2 fixation (King, 2003). A *coxL* gene was found in the *de novo* assembled transcripts affiliated to *Stappia* spp., but not RuBisCO. Sequences analogue to transporters for simple and multiple sugars such as xylose and fructose, and acetate were found (Documents S5–S7), which suggest that the *Stappia* bacteria fed on the biofilm material for carbon sources. Combined with the isolation of denitrifying isolates affiliated to *Marinobacter* spp. and *Paracoccus* spp., these results suggest that the biofilm has the potential to adapt to heterotrophic non-methylotrophic environments.

The proportion of 16S rRNA gene sequences associated to *Bacteroidetes* in the OB and in the eight biofilm cultures ranged from 0.2% to 4.9%, and several genera of this phylum were identified in the three metatranscriptomes. Significant proportions of bacteria affiliated to the *Bacteroidetes* phylum were also found in other methanol-fed denitrification systems. For instance, 29% of cloned 16S rRNA gene sequences were affiliated to *Bacteroidetes* in an acclimated activated sludge in a methanol-fed anoxic denitrification process treating a synthetic wastewater with 4% NaCl (Osaka et al., 2008). Isolates affiliated to the *Bacteroidetes* *Aequorivita* spp. and *Winogradskyella* spp. were isolated from the Ref300N-23C biofilm cultures. None of these isolates, however, could sustain growth under denitrifying conditions. Although denitrification genes affiliated to *Bacteroidetes* genera were found in *de novo* assembled transcripts, genes encoding all four denitrification reductases were not found to any of them. These results suggest that *Bacteroidetes* are not involved in denitrification, although they may be involved in some steps of the denitrification pathway.

The metatranscriptomic data provided some insights of specific metabolisms in *H. nitratorans* strain NL23 and *M. nitratorans* strain GP59 that were regulated in the biofilm environment. In absence of strain NL23 in the Ref300C-23C and the 900N-30C biofilm cultures, the *nirK* gene of strain GP59 was upregulated by 5–10 times compared to the planktonic pure cultures that were also cultivated under denitrifying conditions. On the contrary, the relative transcript levels of this gene did not change between the 0% NaCl biofilm cultures and the planktonic pure cultures, while the relative transcript levels of the NL23 *nirK* were 49 times higher in the 0% NaCl biofilm cultures. These results suggest coordination in the expression of *nirK* between these two strains in the 0% NaCl biofilm cultures.

The gene clusters encoding the three other denitrification reductases (*nap*, *nor*, *nos*) in strain NL23 showed higher relative transcript levels in the 0% NaCl biofilm cultures. *napGH* was however down regulated in these biofilm cultures. *napGH* is located in a separate chromosomal region than the *napABCDEF* operon. NapGH and NapC have redundant function of transferring electrons to NapB across the membrane. It was proposed that NapC transfers electrons from the menaquinol, whereas NapGH do it from ubiquinol (Simon, 2011). The physiological consequence of *napGH* transcript decrease in the biofilm is unknown. Observations on *napEDABC* found in the denitrifier *Shewanella denitrificans* OS217, and *napDAGHB* in the respiratory NO_3^- ammonifier *Shewanella oneidensis* MR-1 suggest that NapGH is more involved in the ammonification system (Simpson, Richardson & Codd, 2010). Despite the denitrifying conditions applied in both types of cultures, the biofilm environment has induced strong up regulation of denitrification genes in *H. nitratorans* strain NL23. This may be in response to the rapid processing of NO_3^- by *M. nitratorans* strain JAM1/GP59 (Mauffrey, Martineau & Villemur, 2015) that could generate rapidly high level of NO_2^- , which is toxic for strain NL23.

CONCLUSION

The OB taken from the Biodome denitrification system underwent substantial changes in its bacterial community when subjected to environmental changes. Cultivating the OB in the homemade ASW medium with different formulations (varying NaCl, NO_3^- and methanol concentrations, and temperature) or in the commercial IO medium showed much higher proportions of *Methylophaga* spp. in these biofilm cultures compared to the OB. These results concur with the growth of *M. nitratorans* strain GP59 in these cultures. The population of *Hyphomicrobium* spp. showed a more complex trend. It was at very low levels in the biofilm cultures cultivated in ASW at 2.75% NaCl, but persisted in the biofilm cultures cultivated in ASW at low NaCl concentration, and also cultivated in the IO medium. Other denitrifiers affiliated to *Marinobacter* spp. and *Paracoccus* spp. were isolated from the biofilm cultures. Moreover, metatranscriptomic analysis revealed that denitrifying bacteria affiliated to *Stappia* spp. were metabolically active in the biofilm cultures. The biofilm environment has favored the upregulation of the denitrification pathway in *M. nitratorans* strain GP59 and *H. nitratorans* strain NL23 compared to planktonic pure cultures, despite the facts that these two types of cultures

were grown under denitrifying conditions. All these results demonstrated the dynamics and the plasticity of the denitrifying biofilm to sustain environmental changes and illustrate a comprehensive picture of the microbial community of the biofilm and its adaptation to these changes. This could benefit in the development of optimal denitrifying bioprocesses under marine conditions. For instance, the fact that non-methylotrophic, denitrifying bacteria are present in the biofilm could suggest adaptation of the denitrification process to another source of carbon such as ethanol or acetate.

ACKNOWLEDGEMENTS

We thank Karla Vasquez for her technical assistance.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This research was supported by a grant to Richard Villemur from the Natural Sciences and Engineering Research Council of Canada # RGPIN-2016-06061. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Natural Sciences and Engineering Research Council of Canada: # RGPIN-2016-06061.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Richard Villemur conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Geneviève Payette and Valérie Geoffroy conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Florian Mauffrey analyzed the data, authored or reviewed drafts of the paper, approved the final draft.
- Christine Martineau conceived and designed the experiments, authored or reviewed drafts of the paper, approved the final draft.

DNA Deposition

The following information was supplied regarding the deposition of DNA sequences:

16S rRNA sequence reads are available in the GenBank Sequence Read Archive (SRA) under the accession number [PRJNA524642](#).

The 16S rRNA gene sequences are available in GenBank under the accession numbers [MK571459–MK571476](#).

RNAseq reads from the planktonic pure cultures and the biofilm cultures are available in the SRA under the accession number [PRJNA525230](#).

De novo assembled transcripts are available in SRA under the accession number [PRJNA525230](#).

Annotations of the *de novo* assembled transcripts are available at the Joint Genomic Institute (<https://img.jgi.doe.gov/cgi-bin/m/main.cgi>) under the GOLD Analysis Project Id: Ga0307915, Ga0307877, Ga0307760.

Data Availability

The following information was supplied regarding data availability:

Raw data is available in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.7467#supplemental-information>.

REFERENCES

- Atlas RM. 1993. In: Parks LC, ed. *Handbook of microbiological media*. Boca Raton: CRC press, 543.
- Auclair J, Lépine F, Parent S, Villemur R. 2010. Dissimilatory reduction of nitrate in seawater by a *Methylophaga* strain containing two highly divergent *narG* sequences. *ISME Journal* 4:1302–1313 DOI [10.1038/ismej.2010.47](#).
- Auclair J, Parent S, Villemur R. 2012. Functional diversity in the denitrifying biofilm of the methanol-fed marine denitrification system at the Montreal Biodome. *Microbial Ecology* 63:726–735 DOI [10.1007/s00248-011-9960-2](#).
- Baytshtok V, Kim S, Yu R, Park H, Chandran K. 2008. Molecular and biokinetic characterization of methylotrophic denitrification using nitrate and nitrite as terminal electron acceptors. *Water Science and Technology* 58:359–365 DOI [10.2166/wst.2008.391](#).
- Cabrol L, Malhautier L. 2011. Integrating microbial ecology in bioprocess understanding: the case of gas biofiltration. *Applied Microbiology and Biotechnology* 90:837–849 DOI [10.1007/s00253-011-3191-9](#).
- Chistoserdova L, Chen SW, Lapidus A, Lidstrom ME. 2003. Methylotrophy in *Methylobacterium extorquens* AM1 from a genomic point of view. *Journal of Bacteriology* 185:2980–2987 DOI [10.1128/Jb.185.10.2980-2987.2003](#).
- Chistoserdova L, Laukel M, Portais JC, Vorholt JA, Lidstrom ME. 2004. Multiple formate dehydrogenase enzymes in the facultative methylotroph *Methylobacterium extorquens* AM1 are dispensable for growth on methanol. *Journal of Bacteriology* 186:22–28 DOI [10.1128/JB.186.1.22-28.2004](#).
- Clark ME, He ZL, Redding AM, Joachimiak MP, Keasling JD, Zhou JZZ, Arkin AP, Mukhopadhyay A, Fields MW. 2012. Transcriptomic and proteomic analyses of *Desulfovibrio vulgaris* biofilms: carbon and energy flow contribute to the distinct biofilm growth state. *BMC Genomics* 13:138 DOI [10.1186/1471-2164-13-138](#).

- Cole JR, Wang Q, Fish JA, Chai B, McGarrell DM, Sun Y, Brown CT, Porras-Alfaro A, Kuske CR, Tiedje JM. 2014. Ribosomal Database Project: data and tools for high throughput rRNA analysis. *Nucleic Acids Research* 42:D633–D642 DOI 10.1093/nar/gkt1244.
- Edgar RC. 2013. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nature Methods* 10:996–998 DOI 10.1038/Nmeth.2604.
- Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. 2011. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics* 27:2194–2200 DOI 10.1093/bioinformatics/btr381.
- Geoffroy V, Payette G, Mauffrey F, Lestin L, Constant P, Villemur R. 2018. Strain-level genetic diversity of *Methylophaga nitratreducenticrescens* confers plasticity to denitrification capacity in a methylotrophic marine denitrifying biofilm. *PeerJ* 6:e4679 DOI 10.7717/peerj.4679.
- Ginige MP, Hugenholtz P, Daims H, Wagner M, Keller J, Blackall LL. 2004. Use of stable-isotope probing, full-cycle rRNA analysis, and fluorescence in situ hybridization-microautoradiography to study a methanol-fed denitrifying microbial community. *Applied and Environmental Microbiology* 70:588–596 DOI 10.1128/Aem.70.1.588-596.2004.
- Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, Adiconis X, Fan L, Raychowdhury R, Zeng Q, Chen Z, Mauceli E, Hacohen N, Gnirke A, Rhind N, Palma Fdi, Birren BW, Nusbaum C, Lindblad-Toh K, Friedman N, Regev A. 2011. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature Biotechnology* 29:644–652 DOI 10.1038/nbt.1883.
- Grob C, Taubert M, Howat AM, Burns OJ, Dixon JL, Richnow HH, Jehmlich N, Von Bergen M, Chen Y, Murrell JC. 2015. Combining metagenomics with metaproteomics and stable isotope probing reveals metabolic pathways used by a naturally occurring marine methylotroph. *Environmental Microbiology* 17:4007–4018 DOI 10.1111/1462-2920.12935.
- Hallin S, Throback IN, Dicksved J, Pell M. 2006. Metabolic profiles and genetic diversity of denitrifying communities in activated sludge after addition of methanol or ethanol. *Applied and Environmental Microbiology* 72:5445–5452 DOI 10.1128/AEM.00809-06.
- Huang C, Zhao YK, Li ZL, Yuan Y, Chen C, Tan WB, Gao S, Gao LF, Zhou JZ, Wang AJ. 2015. Enhanced elementary sulfur recovery with sequential sulfate-reducing, denitrifying sulfide-oxidizing processes in a cylindrical-type anaerobic baffled reactor. *Bioresour Technol* 192:478–485 DOI 10.1016/j.biortech.2015.04.103.
- King GM. 2003. Molecular and culture-based analyses of aerobic carbon monoxide oxidizer diversity. *Applied and Environmental Microbiology* 69:7257–7265 DOI 10.1128/AEM.69.12.7257-7265.2003.
- Kraft B, Strous M, Tegetmeyer HE. 2011. Microbial nitrate respiration—genes, enzymes and environmental distribution. *Journal of Biotechnology* 155:104–117 DOI 10.1016/j.jbiotec.2010.12.025.

- Labbé N, Juteau P, Parent S, Villemur R. 2003.** Bacterial diversity in a marine methanol-fed denitrification reactor at the Montreal Biodome Canada. *Microbial Ecology* 46:12–21 DOI [10.1007/s00248-002-1056-6](https://doi.org/10.1007/s00248-002-1056-6).
- Labbé N, Laurin V, Juteau P, Parent S, Villemur R. 2007.** Microbiological community structure of the biofilm of a methanol-fed, marine denitrification system, and identification of the methanol-utilizing microorganisms. *Microbial Ecology* 53:621–630 DOI [10.1007/s00248-006-9168-z](https://doi.org/10.1007/s00248-006-9168-z).
- Labbé N, Parent S, Villemur R. 2004.** *Nitratireductor aquibiodomus* gen. nov., sp nov., a novel alpha-proteobacterium from the marine denitrification system of the Montreal Biodome (Canada). *International Journal of Systematic and Evolutionary Microbiology* 54:269–273 DOI [10.1099/ijs.0.02793-0](https://doi.org/10.1099/ijs.0.02793-0).
- Li B, Dewey CN. 2011.** RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. *BMC Bioinformatics* 12:323 DOI [10.1186/1471-2105-12-323](https://doi.org/10.1186/1471-2105-12-323).
- Lu HJ, Chandran K, Stensel D. 2014.** Microbial ecology of denitrification in biological wastewater treatment. *Water Research* 64:237–254 DOI [10.1016/j.watres.2014.06.042](https://doi.org/10.1016/j.watres.2014.06.042).
- Martineau C, Mauffrey F, Villemur R. 2015.** Comparative analysis of denitrifying activities of *Hyphomicrobium nitrativorans*, *Hyphomicrobium denitrificans*, and *Hyphomicrobium zavarzinii*. *Applied and Environmental Microbiology* 81:5003–5014 DOI [10.1128/Aem.00848-15](https://doi.org/10.1128/Aem.00848-15).
- Martineau C, Villeneuve C, Mauffrey F, Villemur R. 2013a.** Complete genome sequence of *Hyphomicrobium nitrativorans* strain NL23, a denitrifying bacterium isolated from biofilm of a methanol-fed denitrification system treating seawater at the Montreal Biodome. *Genome Announc* 2:e01165–13 DOI [10.1128/genomeA.01165-13](https://doi.org/10.1128/genomeA.01165-13).
- Martineau C, Villeneuve C, Mauffrey F, Villemur R. 2013b.** *Hyphomicrobium nitrativorans* sp. nov., isolated from the biofilm of a methanol-fed denitrification system treating seawater at the Montreal Biodome. *International Journal of Systematic and Evolutionary Microbiology* 63:3777–3781 DOI [10.1099/ijs.0.048124-0](https://doi.org/10.1099/ijs.0.048124-0).
- Mauffrey F, Cucaita A, Constant P, Villemur R. 2017.** Denitrifying metabolism of the methylotrophic marine bacterium *Methylophaga nitratireducenticrescens* strain JAM1. *PeerJ* 5:e4098 DOI [10.7717/Peerj.4098](https://doi.org/10.7717/Peerj.4098).
- Mauffrey F, Martineau C, Villemur R. 2015.** Importance of the two dissimilatory (Nar) nitrate reductases in the growth and nitrate reduction of the methylotrophic marine bacterium *Methylophaga nitratireducenticrescens* JAM1. *Frontiers in Microbiology* 6:Article 1475 DOI [10.3389/fmicb.2015.01475](https://doi.org/10.3389/fmicb.2015.01475).
- Metsalu T, Vilo J. 2015.** ClustVis: a web tool for visualizing clustering of multivariate data using Principal Component Analysis and heatmap. *Nucleic Acids Research* 43:W566–W570 DOI [10.1093/nar/gkv468](https://doi.org/10.1093/nar/gkv468).
- Neef A, Zaglauer A, Meier H, Amann R, Lemmer H, Schleifer KH. 1996.** Population analysis in a denitrifying sand filter: conventional and in situ identification of *Paracoccus* spp. in methanol-fed biofilms. *Applied and Environmental Microbiology* 62:4329–4339.

- Osaka T, Shirotani K, Yoshie S, Tsuneda S. 2008.** Effects of carbon source on denitrification efficiency and microbial community structure in a saline wastewater treatment process. *Water Research* **42**:3709–3718 DOI [10.1016/j.watres.2008.06.007](https://doi.org/10.1016/j.watres.2008.06.007).
- Osaka T, Yoshie S, Tsuneda S, Hirata A, Iwami N, Inamori Y. 2006.** Identification of acetate- or methanol-assimilating bacteria under nitrate-reducing conditions by stable-isotope probing. *Microbial Ecology* **52**:253–266 DOI [10.1007/s00248-006-9071-7](https://doi.org/10.1007/s00248-006-9071-7).
- Payette G, Geoffroy V, Martineau C, Villemur R. 2019.** Dynamics of a methanol-fed marine denitrifying biofilm: 1-Impact of environmental changes on the denitrification and the co-occurrence of *Methylophaga nitratreducentis* and *Hyphomicrobium nitrativorans*. *PeerJ* **7**:e7497 DOI [10.7717/peerj.7497](https://doi.org/10.7717/peerj.7497).
- Peck V, Quiza L, Buffet JP, Khdhiri M, Durand AA, Paquette A, Thiffault N, Messier C, Beaulieu N, Guertin C, Constant P. 2016.** Towards the development of multifunctional molecular indicators combining soil biogeochemical and microbiological variables to predict the ecological integrity of silvicultural practices. *Microbial Biotechnology* **9**:316–329 DOI [10.1111/1751-7915.12348](https://doi.org/10.1111/1751-7915.12348).
- Peyraud R, Kiefer P, Christen P, Massou S, Portais JC, Vorholt JA. 2009.** Demonstration of the ethylmalonyl-CoA pathway by using C-13 metabolomics. *Proceedings of the National Academy of Sciences of the United States of America* **106**:4846–4851 DOI [10.1073/pnas.0810932106](https://doi.org/10.1073/pnas.0810932106).
- Philippot L, Hojberg O. 1999.** Dissimilatory nitrate reductases in bacteria. *Biochimica Et Biophysica Acta-Gene Structure and Expression* **1446**:1–23 DOI [10.1016/S0167-4781\(99\)00072-X](https://doi.org/10.1016/S0167-4781(99)00072-X).
- Resch A, Rosenstein R, Nerz C, Gotz F. 2005.** Differential gene expression profiling of *Staphylococcus aureus* cultivated under biofilm and planktonic conditions. *Applied and Environmental Microbiology* **71**:2663–2676 DOI [10.1128/Aem.71.5.2663-2676.2005](https://doi.org/10.1128/Aem.71.5.2663-2676.2005).
- Richardson D, Berks B, Russell D, Spiro S, Taylor C. 2001.** Functional, biochemical and genetic diversity of prokaryotic nitrate reductases. *Cellular and Molecular Life Sciences* **58**:165–178 DOI [10.1007/PL00000845](https://doi.org/10.1007/PL00000845).
- Rissanen AJ, Ojala A, Dernjatin M, Jaakkola J, Tiirola M. 2016.** *Methylophaga* and *Hyphomicrobium* can be used as target genera in monitoring saline water methanol-utilizing denitrification. *Journal of Industrial Microbiology and Biotechnology* **43**:1647–1657 DOI [10.1007/s10295-016-1839-2](https://doi.org/10.1007/s10295-016-1839-2).
- Rissanen AJ, Ojala A, Fred T, Toivonen J, Tiirola M. 2017.** *Methylophilaceae* and *Hyphomicrobium* as target taxonomic groups in monitoring the function of methanol-fed denitrification biofilters in municipal wastewater treatment plants. *Journal of Industrial Microbiology and Biotechnology* **44**:35–47 DOI [10.1007/s10295-016-1860-5](https://doi.org/10.1007/s10295-016-1860-5).
- Roder HL, Sorensen SJ, Burmolle M. 2016.** Studying bacterial multispecies biofilms: where to start? *Trends in Microbiology* **24**:503–513 DOI [10.1016/j.tim.2016.02.019](https://doi.org/10.1016/j.tim.2016.02.019).
- Romanenko LA, Tanaka N, Frolova GM, Mikhailov VV. 2010.** *Marinicella litoralis* gen. nov., sp. nov., a gammaproteobacterium isolated from coastal seawater.

- International Journal of Systematic and Evolutionary Microbiology* **60**:1613–1619
DOI 10.1099/ij.s.0.016147-0.
- Salta M, Wharton JA, Blache Y, Stokes KR, Briand JF. 2013.** Marine biofilms on artificial surfaces: structure and dynamics. *Environmental Microbiology* **15**:2879–2893
DOI 10.1111/1462-2920.12186.
- Simon J. 2011.** Organization of respiratory electron transport chains in nitrate-reducing and nitrifying bacteria. In: Moir JWB, ed. *Nitrogen cycling in bacteria Molecular analysis*. Norfolk: Caister academic press, 39–58.
- Simpson PJ, Richardson DJ, Codd R. 2010.** The periplasmic nitrate reductase in *Shewanella*: the resolution, distribution and functional implications of two NAP isoforms, NapEDABC and NapDAGHB. *Microbiology* **156**:302–312
DOI 10.1099/mic.0.034421-0.
- Sun Y, Shen D, Zhou X, Shi N, Tian Y. 2016.** Microbial diversity and community structure of denitrifying biological filters operated with different carbon sources. *Springerplus* **5**:Article 1752 DOI 10.1186/s40064-016-3451-3.
- Tan CH, Lee KW, Burmolle M, Kjelleberg S, Rice SA. 2017.** All together now: experimental multispecies biofilm model systems. *Environmental Microbiology* **19**:42–53
DOI 10.1111/1462-2920.13594.
- Tarazona S, Garcia-Alcalde F, Dopazo J, Ferrer A, Conesa A. 2011.** Differential expression in RNA-seq: a matter of depth. *Genome Research* **21**:2213–2223
DOI 10.1101/gr.124321.111.
- Team RC. 2015.** *A language and environment for statistical computing*. Vienna: R Foundation for Statistical Computing. Available at <https://www.R-project.org/>.
- Villeneuve C, Martineau C, Mauffrey F, Villemur R. 2012.** Complete genome sequences of *Methylophaga* sp. strain JAM1 and *Methylophaga* sp. strain JAM7. *Journal of Bacteriology* **194**:4126–4127 DOI 10.1128/Jb.00726-12.
- Villeneuve C, Martineau C, Mauffrey F, Villemur R. 2013.** *Methylophaga nitratreducens* sp. nov. and *Methylophaga frappieri* sp. nov. isolated from the biofilm of the methanol-fed denitrification system treating the seawater at the Montreal Biodome. *International Journal of Systematic and Evolutionary Microbiology* **63**:2216–2222
DOI 10.1099/Ijs.0.044545-0.
- Wang XQ, Li CM, Dunlap CA, Rooney AP, Du ZJ. 2018.** *Marinicella sediminis* sp nov., isolated from marine sediment. *International Journal of Systematic and Evolutionary Microbiology* **68**:2335–2339 DOI 10.1099/ijsem.0.002839.
- Weber CF, King GM. 2007.** Physiological, ecological, and phylogenetic characterization of *Stappia*, a marine CO-oxidizing bacterial genus. *Applied and Environmental Microbiology* **73**:1266–1276 DOI 10.1128/AEM.01724-06.
- Wright ES, Yilmaz LS, Noguera DR. 2012.** DECIPHER, a search-based approach to chimera identification for 16S rRNA sequences. *Applied and Environmental Microbiology* **78**:717–725 DOI 10.1128/AEM.06516-11.
- Yoshie S, Ogawa T, Makino H, Hirosawa H, Tsuneda S, Hirata A. 2006.** Characteristics of bacteria showing high denitrification activity in saline wastewater. *Letters in Applied Microbiology* **42**:277–283 DOI 10.1111/j.1472-765X.2005.01839.x.